JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
Hands-On Workshop on
"Cytogenetics, Cell & Molecular Biology Techniques"
June 18th to 24th 2022
Organized Under DST STUTI Scheme

FEEDBACK

Evaluation is to be done to assess

1. **Quality of presentation:** In evaluating quality, please take into account speed and style of delivery of speech and quality of slides, besides other characteristics that appeal to you.
2. **Content:** In evaluating the content, please consider whether the material presented was factual, relevant, and up-to-date.
3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	4	4	5
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	4	4	4
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	3	3	3
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	5	5	5
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	4	4	4
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	4	4	4
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood	Ms. Anju Srinivas	5	5	5
	2) ARMS PCR for mutation analysis	Ms. Sumaiya Kalaigar	5	5	5
	3) DNA isolation from bacteria	Mrs. Nishitha R Kumar	5	5	5
	4) 16SrDNA PCR	Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	5	5	5
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	5	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas			
	2) cDNA synthesis	Ms. Vaibhavishree M			
	3) Quantitative real time PCR	Ms. Deepthi V			
	4) Data analysis	Ms. Supriya B			
		Mr. Anshu Kumar Yadav			

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? Yes/No
   b. Have you received handouts of all the sessions? Yes/No
   c. Did the sessions help in improvement of your understanding? Yes/No
   d. Was there adequate time for discussion? Yes/No
   e. Do you think there is need for regular workshops of this type in future?
      Yes/No If yes, how often? _____
   f. Any more techniques you can suggest which can be included in the workshop
      _____
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant? No

j. Suggestions for future workshop
More on Individual hands on trial

Date: 24/06/2022

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3.	RNA interference and its applications	Dr. Prasanna Kumar S,	5	5	5
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	5	5	5
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	5	5	5
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	+5	+5	+5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	5	5
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	5	5	5
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	++5	++5	++5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood 2) ARMS PCR for mutation analysis	Ms. Anju Srinivas	5	5	5
		Ms. Sumaiya Kalaigar	5	5	5
	3) DNA isolation from bacteria 4) 16SrDNA PCR	Mrs. Nishitha R Kumar	5	5	5
		Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	4	4	4
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	5	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check 2) cDNA synthesis 3) Quantitative real time PCR 4) Data analysis	Ms. Anju Srinivas	5	5	5
		Ms. Vaibhavishree M	5	5	5
		Ms. Deepthi V	5	5	5
		Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav	5	5	5

4. Evaluation of the program as a whole:

   a. Did the topics selected cover all important aspects of the specialty? Yes/No

   b. Have you received handouts of all the sessions? Yes/No

   c. Did the sessions help in improvement of your understanding? Yes/No

   d. Was there adequate time for discussion? Yes/No

   e. Do you think there is need for regular workshops of this type in future?

      Yes/No If yes, how often? _____

   f. Any more techniques you can suggest which can be included in the workshop

      Yes, Just Hands practice for each session

   g. How did you get the information regarding this workshop?

      i. By circulars to your institution

      ii. Invitation by organizers

      iii. Internet / Institution website/ social media

      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	5 ✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

NO, I would like to suggest one thing that Tamil Session like explanation slowly, explain it more. from Basic & increased the workshop. period like 15 days to 20 days.

j. Suggestions for future workshop

Time mangment is very good. All staffs, Speakers gave good information.

Date: 24/06/2022

Name and Signature
(Supriya dhotre.)

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2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	4	4
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	3	3	3
4.	Basics of Primer Designing	Dr. Akila Prashant,	4	4	4
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	4
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	3	3	4
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	4	4	4
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	4	4	4
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	4	3	4
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	4	4	4
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	5	4

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B			
		Mr. Manju HC	5	4	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood	Ms. Anju Srinivas			
	2) ARMS PCR for mutation analysis	Ms. Sumaiya Kalaigar	5	5	5
	3) DNA isolation from bacteria	Mrs. Nishitha R Kumar	3	4	4
	4) 16SrDNA PCR	Mr. Mohammed Kaleem Ullah			
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	3	4	4
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	4	4	4
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas	5	5	5
	2) cDNA synthesis	Ms. Vaibhavishree M	5	5	5
	3) Quantitative real time PCR	Ms. Deepthi V	4		
	4) Data analysis	Ms. Supriya B	5	5	4
		Mr. Anshu Kumar Yadav	4	5	4

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? ✓Yes/No
   b. Have you received handouts of all the sessions? ✓Yes/No
   c. Did the sessions help in improvement of your understanding? ✓Yes/No
   d. Was there adequate time for discussion? ✓Yes/No
   e. Do you think there is need for regular workshops of this type in future?
      ✓Yes/No If yes, how often? Twice or Thrice in a MONTH
   f. Any more techniques you can suggest which can be included in the workshop
      CRISPR-Cas 9 & HPLC
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. ✓ Internet / Institution website/ social media
      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

— No —

j. Suggestions for future workshop

Workshop's related to CRISPR - Cas 9.

Date: 24/06/2022

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2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	4	3	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	2	4	5
4.	Basics of Primer Designing	Dr. Akila Prashant,	4	2	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	4	3	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	3	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	4	2	2
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	4	3	5
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	4	2	3
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	3	3
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	3	4
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	2	3	2
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	4	4	4
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	4	5	3
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	4	4	3

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	3	3	3
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood 2) ARMS PCR for mutation analysis 3) DNA isolation from bacteria 4) 16SrDNA PCR	Ms. Anju Srinivas →	4	5	5
		Ms. Sumaiya Kalaigar →	5	5	5
		Mrs. Nishitha R Kumar →	0	2	2
		Mr. Mohammed Kaleem Ullah	5	4	4
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing 4) Collection of protein lysates & Western blot	Mr. Adel Mohammed Saleh. Good work considering he is 'international'	4	3	4
		Mr. Anshu Kumar Yadav	3	4	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check 2) cDNA synthesis 3) Quantitative real time PCR 4) Data analysis	Ms. Anju Srinivas	4	4	4
		Ms. Vaibhavishree M	3	4	4
		Ms. Deepthi V			
		Ms. Supriya B	4	4	4
		Mr. Anshu Kumar Yadav	3	4	5

4. Evaluation of the program as a whole:

   a. Did the topics selected cover all important aspects of the specialty? Yes/No

   b. Have you received handouts of all the sessions? Yes/No

   c. Did the sessions help in improvement of your understanding? Yes/No

   d. Was there adequate time for discussion? Yes/No

   e. Do you think there is need for regular workshops of this type in future?

      Yes/No If yes, how often? Quarterly once

   f. Any more techniques you can suggest which can be included in the workshop

      Cell Based Assays.

   g. How did you get the information regarding this workshop?

      i. By circulars to your institution

      ii. Invitation by organizers

      iii. Internet / Institution website/ social media

      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

None

j. Suggestions for future workshop

A more Hands-on component would be Beneficial.
Enjoyed thoroughly! Good work!

Date: 24/06/2022

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20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas	5	5	5
	2) cDNA synthesis	Ms. Vaibhavishree M	5	5	5
	3) Quantitative real time PCR	Ms. Deepthi V	5	5	5
	4) Data analysis	Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav	5	5	5

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   a. Did the topics selected cover all important aspects of the specialty? ~~Yes~~/No
   b. Have you received handouts of all the sessions? Yes/No
   c. Did the sessions help in improvement of your understanding? Yes/No
   d. Was there adequate time for discussion? Yes/No
   e. Do you think there is need for regular workshops of this type in future?
      ~~Yes~~/No If yes, how often? 2-3 times in a year
   f. Any more techniques you can suggest which can be included in the workshop
      —
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

None

j. Suggestions for future workshop

Inclusion of other techniques and teaching basics of DNA/protein/mRNA isolation with practical.

Date: 24/06/2022

Name and Signature

Dr. Sujit Kumar Sinha

Sujit Kumar Sinha

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4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	4	3	3
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	3	3	2
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	5	5	4
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
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11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	4	5
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	5	5	2
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
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		Mr. Manju HC	5	5	5
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		Ms. Sumaiya Kalaigar	5	5	5
	3) DNA isolation from bacteria 4) 16SrDNA PCR	Mrs. Nishitha R Kumar			
		Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	3	3	3
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	5	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas	5	5	5
	2) cDNA synthesis	Ms. Vaibhavishree M			
	3) Quantitative real time PCR	Ms. Deepthi V			
	4) Data analysis	Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav	5	5	5

4. Evaluation of the program as a whole:

   a. Did the topics selected cover all important aspects of the specialty? Yes/No

   b. Have you received handouts of all the sessions? Yes/No

   c. Did the sessions help in improvement of your understanding? Yes/No

   d. Was there adequate time for discussion? Yes/No

   e. Do you think there is need for regular workshops of this type in future?

      Yes/No If yes, how often? Quarterly

   f. Any more techniques you can suggest which can be included in the workshop

      MS, HPLC, TEM.

   g. How did you get the information regarding this workshop?

      i. By circulars to your institution

      ii. Invitation by organizers

      iii. Internet / Institution website/ social media

      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant? Nil.

j. Suggestions for future workshop - More hands on if possible, newer techniques, single cell genomics & metaphase hands on or demo ~~wise~~

Date: 24/06/2022

Name and Signature

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
Hands-On Workshop on
"Cytogenetics, Cell & Molecular Biology Techniques"
June 18th to 24th 2022
Organized Under DST STUTI Scheme

FEEDBACK

Evaluation is to be done to assess

1. **Quality of presentation:** In evaluating quality, please take into account speed and style of delivery of speech and quality of slides, besides other characteristics that appeal to you.
2. **Content:** In evaluating the content, please consider whether the material presented was factual, relevant, and up-to-date.
3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	4	5	5
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	4	4
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	4	4	4
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	4	4	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	4	3	4
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	5	4	5
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	4	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	4	5	4
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	5	5	5
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	4	4	4
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	4	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	4	4	4

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood	Ms. Anju Srinivas	5	5	5
	2) ARMS PCR for mutation analysis	Ms. Sumaiya Kalaigar	5	4	5
	3) DNA isolation from bacteria	Mrs. Nishitha R Kumar	5	4	5
	4) 16SrDNA PCR	Mr. Mohammed Kaleem Ullah	5	5	5.
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	5	5	5
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	5+	5+	5+
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas	5	5	5
	2) cDNA synthesis	Ms. Vaibhavishree M	4	5	5.
	3) Quantitative real time PCR	Ms. Deepthi V	5	4	5.
	4) Data analysis	Ms. Supriya B	5	5	5.
		Mr. Anshu Kumar Yadav	5+	5+	5+

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? ~~Yes~~/No
   b. Have you received handouts of all the sessions? ~~Yes~~/No
   c. Did the sessions help in improvement of your understanding? ~~Yes~~/No
   d. Was there adequate time for discussion? ~~Yes~~/No
   e. Do you think there is need for regular workshops of this type in future?
      ~~Yes~~/No If yes, how often? Once in 6 months.
   f. Any more techniques you can suggest which can be included in the workshop
      _____
   g. How did you get the information regarding this workshop?
      i. ~~✓~~ By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant? No.

j. Suggestions for future workshop

Chandan Gowda. G.O.

Date: 24/06/2022

Name and Signature

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
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FEEDBACK

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2. **Content:** In evaluating the content, please consider whether the material presented was factual, relevant, and up-to-date.
3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade:	5	4	3	2	1
	(Excellent)	(V. Good)	(Good)	(Satisfactory)	(Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	5	5	5
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	4	4
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	5	5	5
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	4	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	4
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	5	4	3
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	5	4	4
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	4	4
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	5	4
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	4	4	4
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood 2) ARMS PCR for mutation analysis	Ms. Anju Srinivas	5	5	5
		Ms. Sumaiya Kalaigar	5	5	5
	3) DNA isolation from bacteria 4) 16SrDNA PCR	Mrs. Nishitha R Kumar	5	5	5
		Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	5	5	5
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	5	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check 2) cDNA synthesis 3) Quantitative real time PCR 4) Data analysis	Ms. Anju Srinivas	5	5	5
		Ms. Vaibhavishree M	5	5	5
		Ms. Deepthi V	5	5	5
		Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav	5	5	5

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? Yes/No
   b. Have you received handouts of all the sessions? Yes/No
   c. Did the sessions help in improvement of your understanding? Yes/No
   d. Was there adequate time for discussion? Yes/No
   e. Do you think there is need for regular workshops of this type in future?
      Yes/No If yes, how often? may be once a year
   f. Any more techniques you can suggest which can be included in the workshop
      Flow cytometry / Mass spectrometry
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food		✓	
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

None

j. Suggestions for future workshop

None

Date: 24/06/2022

K. Meghana

Name and Signature

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
Hands-On Workshop on
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June 18th to 24th 2022
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FEEDBACK

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2. **Content:** In evaluating the content, please consider whether the material presented was factual, relevant, and up-to-date.
3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	3	4	5
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	5	5	5
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	2	1	1
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	4	3	4
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	4	5
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	4	3	4
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood 2) ARMS PCR for mutation analysis	Ms. Anju Srinivas	5	4	4
		Ms. Sumaiya Kalaigar	—	—	—
	3) DNA isolation from bacteria 4) 16SrDNA PCR	Mrs. Nishitha R Kumar	4	3	4
		Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing 4) Collection of protein lysates & Western blot	Mr. Adel Mohammed Saleh	3	4	5
		Mr. Anshu Kumar Yadav	5	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check 2) cDNA synthesis 3) Quantitative real time PCR 4) Data analysis	Ms. Anju Srinivas	5	5	5
		Ms. Vaibhavishree M	4	4	4
		Ms. Deepthi V	5	5	5
		Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav	3	4	5

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? Yes/No
   b. Have you received handouts of all the sessions? Yes/No
   c. Did the sessions help in improvement of your understanding? Yes/No
   d. Was there adequate time for discussion? Yes/No
   e. Do you think there is need for regular workshops of this type in future?
      Yes/No If yes, how often? 2 Times in a Year.
   f. Any more techniques you can suggest which can be included in the workshop
      Cell Freezing,
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

j. Suggestions for future workshop

Continue the Same work

Great job.

Happy to be part of workshop

Date: 24/06/2022

Name and Signature

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
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FEEDBACK

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2. **Content:** In evaluating the content, please consider whether the material presented was factual, relevant, and up-to-date.
3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	4	4	4
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	4	4	4
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	4	5	4
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	4	4	4
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	4	4	4
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	4	4	4
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	4	4	4
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	4	4	4
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	4	4	4
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	4	5	4
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	4	4	4
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood	Ms. Anju Srinivas	4	4	4
	2) ARMS PCR for mutation analysis	Ms. Sumaiya Kalaigar	4	4	4
	3) DNA isolation from bacteria	Mrs. Nishitha R Kumar	4	4	4
	4) 16SrDNA PCR	Mr. Mohammed Kaleem Ullah	4	4	4
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting	Mr. Adel Mohammed Saleh	4	4	4
	3) Subculturing 4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	4	4	4
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas	4	4	4
	2) cDNA synthesis	Ms. Vaibhavishree M	4	4	4
	3) Quantitative real time PCR	Ms. Deepthi V	4	4	4
	4) Data analysis	Ms. Supriya B	4	4	4
		Mr. Anshu Kumar Yadav	4	4	4

4. Evaluation of the program as a whole:

   a. Did the topics selected cover all important aspects of the specialty? ✓ Yes/No

   b. Have you received handouts of all the sessions? ✓ Yes/No

   c. Did the sessions help in improvement of your understanding? ✓ Yes/No

   d. Was there adequate time for discussion? ✓ Yes/No

   e. Do you think there is need for regular workshops of this type in future?

      ✓ Yes/No If yes, how often? Every year (once in year)

   f. Any more techniques you can suggest which can be included in the workshop

      Immunocytochemistry and ELISA

   g. How did you get the information regarding this workshop?

      i. By circulars to your institution

      ~~ii.~~ ✓ Invitation by organizers

      iii. Internet / Institution website/ social media

      ~~iv.~~ ✓ Any other • Through Friends

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

— Nil —

j. Suggestions for future workshop

Kindly organize at least two weeks.

Ambarisha Chabbi

Date: 24/06/2022

Name and Signature

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
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3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

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Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	5	5	3
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	4	4	4
4.	Basics of Primer Designing	Dr. Akila Prashant,	4	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	3	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	3	3	3
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	3	3	3
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	4	4	5
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	3	4	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	4	4	3
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	4	4	4
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	3	3	3
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	4	4	3

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood 2) ARMS PCR for mutation analysis 3) DNA isolation from bacteria 4) 16SrDNA PCR	Ms. Anju Srinivas	5	5	5
		Ms. Sumaiya Kalaigar	na	na	na
		Mrs. Nishitha R Kumar	na	na	na
		Mr. Mohammed Kaleem Ullah	4	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing 4) Collection of protein lysates & Western blot	Mr. Adel Mohammed Saleh	3	3	3
		Mr. Anshu Kumar Yadav	3	4	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check 2) cDNA synthesis 3) Quantitative real time PCR 4) Data analysis	Ms. Anju Srinivas	5	5	5
		Ms. Vaibhavishree M			
		Ms. Deepthi V			
		Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav	4	5	5

4. Evaluation of the program as a whole:

   a. Did the topics selected cover all important aspects of the specialty? Yes/~~No~~

   b. Have you received handouts of all the sessions? Yes/No

   c. Did the sessions help in improvement of your understanding? Yes/~~No~~

   d. Was there adequate time for discussion? Yes/~~No~~

   e. Do you think there is need for regular workshops of this type in future?

      Yes/~~No~~ If yes, how often? Once every year

   f. Any more techniques you can suggest which can be included in the workshop

      Mass spectroscopy, silver staining, Immunohistochemistry, IF

   g. How did you get the information regarding this workshop?

      i. ✓ By circulars to your institution

      ii. Invitation by organizers

      iii. Internet / Institution website/ social media

      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

j. Suggestions for future workshop

Date: 24/06/2022

Name and Signature

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3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

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Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	5	4	4
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	3	4	4
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	4	4	4
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	4	4
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	4	4
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	4	4	4
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	4	4	4
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	4	4
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	4	4	4
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	4	4	4
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	4	4	3
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	4	4
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	4	4	4
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	4	4	4
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	4	4

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood	Ms. Anju Srinivas	5	5	5
	2) ARMS PCR for mutation analysis	Ms. Sumaiya Kalaigar	5	5	5
	3) DNA isolation from bacteria	Mrs. Nishitha R Kumar	4	5	5
	4) 16SrDNA PCR	Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	5	5	5
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	5	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas	5	5	5
	2) cDNA synthesis	Ms. Vaibhavishree M	4	5	5
	3) Quantitative real time PCR	Ms. Deepthi V			
	4) Data analysis	Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav			

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? ✓ Yes/No
   b. Have you received handouts of all the sessions? Yes/No
   c. Did the sessions help in improvement of your understanding? ✓ Yes/No
   d. Was there adequate time for discussion? ✓ Yes/No
   e. Do you think there is need for regular workshops of this type in future?
      ✓ Yes/No If yes, how often? once in a years
   f. Any more techniques you can suggest which can be included in the workshop
      few computational / Bioinformatics techniques and flow cytometry, sequencing etc
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      ✓ iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

None

j. Suggestions for future workshop

—

Date: 24/06/2022

Chaitha

Name and Signature

Sai Chaitha Mullaguri

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
Hands-On Workshop on
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June 18th to 24th 2022
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FEEDBACK

Evaluation is to be done to assess

1. **Quality of presentation:** In evaluating quality, please take into account speed and style of delivery of speech and quality of slides, besides other characteristics that appeal to you.
2. **Content:** In evaluating the content, please consider whether the material presented was factual, relevant, and up-to-date.
3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	Good	4	5
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	Excellent	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	Excellent	5	5
4.	Basics of Primer Designing	Dr. Akila Prashant,	Excellent	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	4	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	3	5	3
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	4	4	4
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	4	4	4
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	4	4	4
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	4	4	5
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	3	4	3
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	4	5	4
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	4	4
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	4	4	4

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	4	4	4
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood	Ms. Anju Srinivas	5	5	5
	2) ARMS PCR for mutation analysis	Ms. Sumaiya Kalaigar	4	4	4
	3) DNA isolation from bacteria	Mrs. Nishitha R Kumar	5	5	5
	4) 16SrDNA PCR	Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	4	4	4
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	5***	5***	5***
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas	5	5	4
	2) cDNA synthesis	Ms. Vaibhavishree M	5	5	5
	3) Quantitative real time PCR	Ms. Deepthi V	3	3	3
	4) Data analysis	Ms. Supriya B	4	4	4
		Mr. Anshu Kumar Yadav	5	5	5

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? ~~Yes/No~~
   b. Have you received handouts of all the sessions? ~~Yes/No~~
   c. Did the sessions help in improvement of your understanding? ~~Yes/No~~
   d. Was there adequate time for discussion? ~~Yes/No~~
   e. Do you think there is need for regular workshops of this type in future?
      ~~Yes/No~~ If yes, how often? 6th monthance.
   f. Any more techniques you can suggest which can be included in the workshop
      _____
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓	✓	
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant? NO

j. Suggestions for future workshop

Kindly conduct the workshop on animal handling and drug dosages & other procedures.

Date: 24/06/2022

Name and Signature

Thankyou.

JSS Academy of Higher Education &Research
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2. **Content:** In evaluating the content, please consider whether the material presented was factual, relevant, and up-to-date.
3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	4	4	4
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	4	4	4
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	5	5	5
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	4	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	3	3	3
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	4	5	5
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	4	4
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	4	4	4
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	4	4	4
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	3	5	3
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	4	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	4	4	4
		Mr. Manju HC	4	4	4
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood 2) ARMS PCR for mutation analysis	Ms. Anju Srinivas	5	5	5
		Ms. Sumaiya Kalaigar	4	4	4
	3) DNA isolation from bacteria 4) 16SrDNA PCR	Mrs. Nishitha R Kumar	4	4	4
		Mr. Mohammed Kaleem Ullah	4	4	4
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	4	4	4
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	4	4	4
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check 2) cDNA synthesis 3) Quantitative real time PCR 4) Data analysis	Ms. Anju Srinivas	5	5	5
		Ms. Vaibhavishree M			
		Ms. Deepthi V			
		Ms. Supriya B	4	5	5
		Mr. Anshu Kumar Yadav			

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? ~~Yes~~/No
   b. Have you received handouts of all the sessions? ~~Yes~~/No
   c. Did the sessions help in improvement of your understanding? ~~Yes~~/No
   d. Was there adequate time for discussion? ~~Yes~~/No
   e. Do you think there is need for regular workshops of this type in future?
      ~~Yes~~/No If yes, how often? _____
   f. Any more techniques you can suggest which can be included in the workshop
      NGS?
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. ~~Any other~~

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

None at all.

j. Suggestions for future workshop

An amazing workshop.

THANK YOU SO MUCH.

Date: 24/06/2022

Name and Signature

Medha.

MEDHA.

Suggestion

1) Divide into smaller groups and have as many stations.

Eg → 5 groups of 6 members each.

so 5 stations + ~~one~~ demonstrator station.

2) Single demonstrator for ~~the whole~~ all the groups. ~~rather than one for~~

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3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

   Evaluation is to be done using the scale outline below: -

   Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	4	5	5
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	4	5	5
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	4	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	2	3	3
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	4	4	5
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	4	4	4
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	3	4	4
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	4	4	4
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	4	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood 2) ARMS PCR for mutation analysis	Ms. Anju Srinivas	5	5	5
		Ms. Sumaiya Kalaigar			
	3) DNA isolation from bacteria 4) 16SrDNA PCR	Mrs. Nishitha R Kumar	4	4	5
		Mr. Mohammed Kaleem Ullah			
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	4	4	4
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	4	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check 2) cDNA synthesis 3) Quantitative real time PCR 4) Data analysis	Ms. Anju Srinivas	5	5	5
		Ms. Vaibhavishree M	4	4	4
		Ms. Deepthi V			
		Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav	4	4	5

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? Yes/No
   b. Have you received handouts of all the sessions? Yes/No
   c. Did the sessions help in improvement of your understanding? Yes/No
   d. Was there adequate time for discussion? Yes/No
   e. Do you think there is need for regular workshops of this type in future?
      Yes/No If yes, how often? One in every 6 months
   f. Any more techniques you can suggest which can be included in the workshop
      Molecular Biology technique – Cloning
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓✓✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

NO

j. Suggestions for future workshop

Wonderful team and excellent group/team work, so pretty much everything was PERFECT!

Kruthika.M

Date: 24/06/2022

Name and Signature

M. KRUTHIKA

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
Hands-On Workshop on
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Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	5	5	4
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	4	5	4
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	4
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	4
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	4	5	5
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	4	5	5
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	5	4
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	5	5	4
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	4	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood 2) ARMS PCR for mutation analysis	Ms. Anju Srinivas	5	5	5
		Ms. Sumaiya Kalaigar	5	4	5
	3) DNA isolation from bacteria 4) 16SrDNA PCR	Mrs. Nishitha R Kumar	5	5	5
		Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting	Mr. Adel Mohammed Saleh	5	4	5
	3) Subculturing 4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	5	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check 2) cDNA synthesis 3) Quantitative real time PCR 4) Data analysis	Ms. Anju Srinivas	5	4	5
		Ms. Vaibhavishree M	5	4	5
		Ms. Deepthi V	5	5	4
		Ms. Supriya B	5	5	4
		Mr. Anshu Kumar Yadav	5	5	5

4. Evaluation of the program as a whole:

   a. Did the topics selected cover all important aspects of the specialty? Yes/No

   b. Have you received handouts of all the sessions? Yes/No

   c. Did the sessions help in improvement of your understanding? Yes/No

   d. Was there adequate time for discussion? Yes/No

   e. Do you think there is need for regular workshops of this type in future?

      Yes/No If yes, how often? Quarterly

   f. Any more techniques you can suggest which can be included in the workshop

      N.A.

   g. How did you get the information regarding this workshop?

      i. By circulars to your institution

      ii. Invitation by organizers

      iii. Internet / Institution website/ social media

      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

NO problems. The workshop was organized very well.

j. Suggestions for future workshop

Any workshop related to biochemistry. Also, on clinical research would be helpful.

Date: 24/06/2022

Name and Signature

AMITH BHARADWAJ. S.A

Amith

JSS Academy of Higher Education &Research
JSS Medical College
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Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	4	5	3
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	4	3
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	5	4	4
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	4	4
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	4
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	4	5	5
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	5	4	3
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10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	5	5
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	5	4	3
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	4	4
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC			
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood	Ms. Anju Srinivas	5	5	5
	2) ARMS PCR for mutation analysis	Ms. Sumaiya Kalaigar			
	3) DNA isolation from bacteria	Mrs. Nishitha R Kumar			
	4) 16SrDNA PCR	Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	5	5	5
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	5	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas	5	4	4
	2) cDNA synthesis	Ms. Vaibhavishree M			
	3) Quantitative real time PCR	Ms. Deepthi V			
	4) Data analysis	Ms. Supriya B	5	4	5
		Mr. Anshu Kumar Yadav	5	5	5

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? Yes/No
   b. Have you received handouts of all the sessions? Yes/No
   c. Did the sessions help in improvement of your understanding? Yes/No
   d. Was there adequate time for discussion? Yes/No
   e. Do you think there is need for regular workshops of this type in future?
      Yes/No If yes, how often? 3 Months
   f. Any more techniques you can suggest which can be included in the workshop
      _____
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

—

j. Suggestions for future workshop

DAMINI. C. S

Name and Signature

Date: 24/06/2022

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
Hands-On Workshop on
"Cytogenetics, Cell & Molecular Biology Techniques"
June 18th to 24th 2022
Organized Under DST STUTI Scheme

FEEDBACK

Evaluation is to be done to assess

1. **Quality of presentation:** In evaluating quality, please take into account speed and style of delivery of speech and quality of slides, besides other characteristics that appeal to you.
2. **Content:** In evaluating the content, please consider whether the material presented was factual, relevant, and up-to-date.
3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	4	5	5
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	4	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	4	4	4
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	4	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	4	4	4
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	3	4	5
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	4	5	5
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	5	5
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	5	5	5
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	4	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	4	4	4

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood 2) ARMS PCR for mutation analysis 3) DNA isolation from bacteria 4) 16SrDNA PCR	Ms. Anju Srinivas	5	5	5
		Ms. Sumaiya Kalaigar	5	5	5
		Mrs. Nishitha R Kumar	5	5	5
		Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing 4) Collection of protein lysates & Western blot	Mr. Adel Mohammed Saleh	4	4	4
		Mr. Anshu Kumar Yadav	5	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check 2) cDNA synthesis 3) Quantitative real time PCR 4) Data analysis	Ms. Anju Srinivas	5	5	5
		Ms. Vaibhavishree M	5	5	5
		Ms. Deepthi V	5	5	5
		Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav			

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? ~~Yes~~/No
   b. Have you received handouts of all the sessions? ~~Yes~~/No
   c. Did the sessions help in improvement of your understanding? ~~Yes~~/No
   d. Was there adequate time for discussion? ~~Yes~~/No
   e. Do you think there is need for regular workshops of this type in future?
      ~~Yes~~/No If yes, how often? _____
   f. Any more techniques you can suggest which can be included in the workshop
      _____
   g. How did you get the information regarding this workshop?
      i. ~~By~~ circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

Satisfied!!!

j. Suggestions for future workshop

KAJAL SHARMA

Name and Signature

Date: 24/06/2022

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
Hands-On Workshop on
"Cytogenetics, Cell & Molecular Biology Techniques"
June 18th to 24th 2022
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FEEDBACK

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3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	5	5	5
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	5	5	5
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	3	3	3
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	5	5	5
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	5	5
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	5	5	5
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC			
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood 2) ARMS PCR for mutation analysis	Ms. Anju Srinivas	5	5	5
		Ms. Sumaiya Kalaigar			
	3) DNA isolation from bacteria 4) 16SrDNA PCR	Mrs. Nishitha R Kumar			
		Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	5	5	5
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	5	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check 2) cDNA synthesis 3) Quantitative real time PCR 4) Data analysis	Ms. Anju Srinivas	5	5	5
		Ms. Vaibhavishree M	5 -	5 -	5 -
		Ms. Deepthi V	5	5	5
		Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav	5	5	5

4. Evaluation of the program as a whole:

   a. Did the topics selected cover all important aspects of the specialty? Yes/No

   b. Have you received handouts of all the sessions? Yes/No

   c. Did the sessions help in improvement of your understanding? Yes/No

   d. Was there adequate time for discussion? Yes/No

   e. Do you think there is need for regular workshops of this type in future?

      Yes/No If yes, how often? Once every 4 months

   f. Any more techniques you can suggest which can be included in the workshop

      LCMS/MS, NABL training

   g. How did you get the information regarding this workshop?

      i. By circulars to your institution

      ii. Invitation by organizers

      iii. Internet / Institution website/ social media

      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

None

j. Suggestions for future workshop

Kindly conduct more such workshop

Nithya Shenoy

Dr. Nithya Shenoy

Date: 24/06/2022

Name and Signature

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
Hands-On Workshop on
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FEEDBACK

Evaluation is to be done to assess

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2. **Content:** In evaluating the content, please consider whether the material presented was factual, relevant, and up-to-date.
3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	Good	Good	Satisfactory
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	3	3	2
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	3	3	2
4.	Basics of Primer Designing	Dr. Akila Prashant,	4	5	5 (yes)
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	3	3	2
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	3	3	2
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	3	3	2
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	3	3	2
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	3	3	2
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	4	4	4
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	4	4	4
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	3	3	3
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	3	4	3
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	3	4	4
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	3	3	3
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	3	3	3

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	3	3	3
		Mr. Manju HC	3	3	3
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood	Ms. Anju Srinivas	3	3	3
	2) ARMS PCR for mutation analysis	Ms. Sumaiya Kalaigar	3	3	3
	3) DNA isolation from bacteria	Mrs. Nishitha R Kumar	3	2	2
	4) 16SrDNA PCR	Mr. Mohammed Kaleem Ullah	2	3	2
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	2	2	2
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	3	3	3
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas	3	3	3
	2) cDNA synthesis	Ms. Vaibhavishree M	3	3	3
	3) Quantitative real time PCR	Ms. Deepthi V	3	2	2
	4) Data analysis	Ms. Supriya B	3	3	3
		Mr. Anshu Kumar Yadav	3	3	3

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? Yes/No
   b. Have you received handouts of all the sessions? Yes/No
   c. Did the sessions help in improvement of your understanding? Yes/No
   d. Was there adequate time for discussion? Yes/No
   e. Do you think there is need for regular workshops of this type in future?
      Yes/No If yes, how often? Continuous improvement and also need to gain knowledge to uptodate technology
   f. Any more techniques you can suggest which can be included in the workshop
      –
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

- No -

j. Suggestions for future workshop

—

Date: 24/06/2022

Name and Signature

(T. N. Ravi)

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
Hands-On Workshop on
"Cytogenetics, Cell & Molecular Biology Techniques"
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FEEDBACK

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3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	4	5	4
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	5	5	5
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	4	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	4
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	5	5	3
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	5	5	5
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	4	3
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	5	5	5
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	4
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	4
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	4
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	4
		Mr. Manju HC	5	5	4
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood	Ms. Anju Srinivas	5	5	4
	2) ARMS PCR for mutation analysis	Ms. Sumaiya Kalaigar	5	5	5
	3) DNA isolation from bacteria	Mrs. Nishitha R Kumar			
	4) 16SrDNA PCR	Mr. Mohammed Kaleem Ullah	5	4	4
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	5	4	5
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav			
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas	5	4	5
	2) cDNA synthesis	Ms. Vaibhavishree M			
	3) Quantitative real time PCR	Ms. Deepthi V			
	4) Data analysis	Ms. Supriya B	4	5	4
		Mr. Anshu Kumar Yadav			

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? Yes/No
   b. Have you received handouts of all the sessions? Yes/No
   c. Did the sessions help in improvement of your understanding? Yes/No
   d. Was there adequate time for discussion? Yes/No
   e. Do you think there is need for regular workshops of this type in future?
      Yes/No If yes, how often? Yearly plz. & updates.
   f. Any more techniques you can suggest which can be included in the workshop
      –
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other – Message from Consultant Dr. B. Abishekh
      Asst. Prof.
      Dept. of Transfusion Medicine
      JIPMER - Puducherry.

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

Nothing to comment, [Well organised].

j. Suggestions for future workshop

- Powerpoint presentations if shared will be helpful.

Date: 24/06/2022

Name and Signature

PSRIRAM
JR/III
MMC19SSR
24/6/2022

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
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Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	3	4	3
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	2	3	3
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	4
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	3	3	2
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	2	2	2
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	3	3	3
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	4	4	3
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	4	4	3
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	2	2	2
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	4
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	3	4	3
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	4
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	3	3	2

quality content usefulness

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B			
		Mr. Manju HC	4	5	3
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood 2) ARMS PCR for mutation analysis	Ms. Anju Srinivas	5	5	5
		Ms. Sumaiya Kalaigar	5	5	5
	3) DNA isolation from bacteria 4) 16SrDNA PCR	Mrs. Nishitha R Kumar	5	5	4
		Mr. Mohammed Kaleem Ullah			
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing 4) Collection of protein lysates & Western blot	Mr. Adel Mohammed Saleh	3	4	3
		Mr. Anshu Kumar Yadav	3	4	3
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check 2) cDNA synthesis 3) Quantitative real time PCR 4) Data analysis	Ms. Anju Srinivas	5	5	5
		Ms. Vaibhavishree M	5	5	5
		Ms. Deepthi V			
		Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav			

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? (Yes)/No
   b. Have you received handouts of all the sessions? (Yes)/No
   c. Did the sessions help in improvement of your understanding? (Yes)/No
   d. Was there adequate time for discussion? (Yes)/No
   e. Do you think there is need for regular workshops of this type in future?
      Yes/No If yes, how often? _____
   f. Any more techniques you can suggest which can be included in the workshop
      _____
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      (iv.) Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	X		
Food	X		
Conduct of workshop	X		

i. Any problems faced by you as a participant?

n/a

j. Suggestions for future workshop

- For ~~During~~ hands-on demonstrations, split up delegates into smaller groups so that there are more opportunities for participation & improve visibility of equipment / lab procedures.

~~more open~~

Date: 24/06/2022

Name and Signature

- overall, the workshop was incredibly well-designed and executed. thank you all for putting in so much time & effort to make this an informative & high-yield experience!

JSS Academy of Higher Education &Research
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Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	5	5	5
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	4	4	4
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	4	4	4
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	4	4	4
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	4	4	4
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	5	5
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	4	4	4
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood 2) ARMS PCR for mutation analysis	Ms. Anju Srinivas	5	5	5
		Ms. Sumaiya Kalaigar	4	4	4
	3) DNA isolation from bacteria 4) 16SrDNA PCR	Mrs. Nishitha R Kumar	4	4	4
		Mr. Mohammed Kaleem Ullah	4	4	4
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	4	4	4
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	5	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check 2) cDNA synthesis 3) Quantitative real time PCR 4) Data analysis	Ms. Anju Srinivas	5	5	5
		Ms. Vaibhavishree M	5	5	5
		Ms. Deepthi V	5	5	5
		Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav	5	5	5

4. Evaluation of the program as a whole:

   a. Did the topics selected cover all important aspects of the specialty? Yes/No

   b. Have you received handouts of all the sessions? Yes/No

   c. Did the sessions help in improvement of your understanding? Yes/No

   d. Was there adequate time for discussion? Yes/No

   e. Do you think there is need for regular workshops of this type in future?

      Yes/No If yes, how often? two times for a year.

   f. Any more techniques you can suggest which can be included in the workshop

      Molecular Docking techniques

   g. How did you get the information regarding this workshop?

      i. By circulars to your institution

      ii. Invitation by organizers

      iii. Internet / Institution website/ social media

      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

NO

j. Suggestions for future workshop

No more suggestions its good.

Narayana . P.

Date: 24/06/2022

Name and Signature

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
Hands-On Workshop on
"Cytogenetics, Cell & Molecular Biology Techniques"
June 18th to 24th 2022
Organized Under DST STUTI Scheme

FEEDBACK

Evaluation is to be done to assess

1. **Quality of presentation:** In evaluating quality, please take into account speed and style of delivery of speech and quality of slides, besides other characteristics that appeal to you.
2. **Content:** In evaluating the content, please consider whether the material presented was factual, relevant, and up-to-date.
3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	5	5	5
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	4	4 1/2	4 1/2
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	4	4 1/2	4 1/2
4.	Basics of Primer Designing	Dr. Akila Prashant,	5+	5+	5+
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	4 1/2	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	4	4	4
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	5	5	4 1/2
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	4 1/2	4 1/2
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	3 1/2	3 1/2	4
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	4	4	4
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	4	4	4
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	4	4	4

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	3½	3½	3
		Mr. Manju HC	5+	5++	5+++
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood 2) ARMS PCR for mutation analysis	Ms. Anju Srinivas	4½	4½	4½
		Ms. Sumaiya Kalaigar	4½	4½	4½
	3) DNA isolation from bacteria 4) 16SrDNA PCR	Mrs. Nishitha R Kumar	4½	4½	4½
		Mr. Mohammed Kaleem Ullah	-4½	4½	4½
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing 4) Collection of protein lysates & Western blot	Mr. Adel Mohammed Saleh			
		Mr. Anshu Kumar Yadav	4½	4½	4½
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check 2) cDNA synthesis 3) Quantitative real time PCR 4) Data analysis	Ms. Anju Srinivas	4	4	4
		Ms. Vaibhavishree M	4	5	5
		Ms. Deepthi V	4	4	4
		Ms. Supriya B	3	3	3
		Mr. Anshu Kumar Yadav	5	4	4

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? Yes/No
   b. Have you received handouts of all the sessions? Yes/No
   c. Did the sessions help in improvement of your understanding? Yes/No
   d. Was there adequate time for discussion? Yes/No
   e. Do you think there is need for regular workshops of this type in future?
      Yes/No If yes, how often? Every 3-6 months
   f. Any more techniques you can suggest which can be included in the workshop
      More Molecular Techniques, & Proteomics
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other
      Friend's Forward messages.

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

No problems

j. Suggestions for future workshop

NA

Date: 24/06/2022

Name and Signature

Nagarjuna D

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
Hands-On Workshop on
"Cytogenetics, Cell & Molecular Biology Techniques"
June 18th to 24th 2022
Organized Under DST STUTI Scheme

FEEDBACK

Evaluation is to be done to assess

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2. **Content:** In evaluating the content, please consider whether the material presented was factual, relevant, and up-to-date.
3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	5	4	3
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	4	3	3
4.	Basics of Primer Designing	Dr. Akila Prashant,	4	4	4
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	4
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	4	4	4
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	3	2	2
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	4	4	4
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	5	5
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	4	4	4
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	4	4	4
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	4	4	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood	Ms. Anju Srinivas	5	5	5
	2) ARMS PCR for mutation analysis	Ms. Sumaiya Kalaigar	5	4	4
	3) DNA isolation from bacteria	Mrs. Nishitha R Kumar	4	4	4
	4) 16SrDNA PCR	Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	5	5	5
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	5	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas	5	5	5
	2) cDNA synthesis	Ms. Vaibhavishree M	5	5	5
	3) Quantitative real time PCR	Ms. Deepthi V	5	5	5
	4) Data analysis	Ms. Supriya B	5	5	4
		Mr. Anshu Kumar Yadav	5	5	4

4. Evaluation of the program as a whole:

   a. Did the topics selected cover all important aspects of the specialty? ~~Yes~~/No

   b. Have you received handouts of all the sessions? ~~Yes~~/No

   c. Did the sessions help in improvement of your understanding? ~~Yes~~/No

   d. Was there adequate time for discussion? ~~Yes~~/No

   e. Do you think there is need for regular workshops of this type in future?

      ~~Yes~~/No If yes, how often? Twice in an year.

   f. Any more techniques you can suggest which can be included in the workshop

      Hands on Mass spectrometry.

   g. How did you get the information regarding this workshop?

      ~~i.~~ By circulars to your institution

      ii. Invitation by organizers

      iii. Internet / Institution website/ social media

      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

There was no problem faced as the participant, it was infact quite smooth conduct.

j. Suggestions for future workshop

ZOYA SHAIKH.

Zoya Shaikh

Date: 24/06/2022

Name and Signature

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
Hands-On Workshop on
"Cytogenetics, Cell & Molecular Biology Techniques"
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3. **Usefulness to you:** In evaluating the usefulness, please consider whether you learned anything now and whether you can use it in the Indian context.

Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	5	5	5
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	5	5	5
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	5	5	5
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	5	5	5
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	5	5
12.	See biology in new ways with single-cell and spatial technologies from 10x Genomics	Dr. Ivonne Petermann	5	5	5
13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC	5	5	5
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood	Ms. Anju Srinivas	5	5	5
	2) ARMS PCR for mutation analysis	Ms. Sumaiya Kalaigar	5	5	5
	3) DNA isolation from bacteria	Mrs. Nishitha R Kumar	5		
	4) 16SrDNA PCR	Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh			
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav	5	5	5
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas	5	5	5
	2) cDNA synthesis	Ms. Vaibhavishree M	5	5	5
	3) Quantitative real time PCR	Ms. Deepthi V	5	5	5
	4) Data analysis	Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav	5	5	5

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? ~~Yes~~/No
   b. Have you received handouts of all the sessions? ~~Yes~~/No
   c. Did the sessions help in improvement of your understanding? ~~Yes~~/No
   d. Was there adequate time for discussion? ~~Yes~~/No
   e. Do you think there is need for regular workshops of this type in future?
      ~~Yes~~/No If yes, how often? _____
   f. Any more techniques you can suggest which can be included in the workshop
      _____
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	5		
Food	5		
Conduct of workshop	5		

i. Any problems faced by you as a participant?

—

j. Suggestions for future workshop

—

M. Mahalakshmi

Date: 24/06/2022

Name and Signature

24/6/2022

JSS Academy of Higher Education &Research
JSS Medical College
Department of Biochemistry& Department of Medical Genetics
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   Evaluation is to be done using the scale outline below: -

Grade: 5 (Excellent) 4 (V. Good) 3 (Good) 2 (Satisfactory) 1 (Not Satisfactory)

Sl No.	Topic	Speaker	Quality	Content	Usefulness
1.	Introduction to Cytogenetics	Dr. P.V.V. Gowri Sairam,	5	5	5
2.	Good Cell Culture Practice	Dr. Divya Prasannakumar,	5	5	5
3.	RNA interference and its applications	Dr. Prasanna Kumar S,	5	5	5
4.	Basics of Primer Designing	Dr. Akila Prashant,	5	5	5
5.	Basics of Real-time PCR	Dr. Rajeshkumar T,	5	5	5
6.	Application of cell-based assays in drug discovery and development	Dr. Subbarao V Madhunapantula,	5	5	5
7.	Metasystems solutions for FISH	Dr. Lakshman Kumar B	5	5	4
8.	Multiplex PCR: Protocol development and applications in medical diagnosis	Dr. D. Devananda,	5	5	5
9.	Molecular Diagnostic tools & Genetic testing in the field of Clinical Medicine	Dr. Pooja Aggarwal,	5	5	5
10.	Most frequent chromosomal abnormalities in genetic disorders	Ms.Supriya B	5	5	5
11.	Online Tools for NGS data analysis	Mr. Mohammed Kaleem Ullah,	5	5	5
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13.	Fundamentals of microscopy	Dr. Vinay Kumar Rao,	5	5	5
14.	Karyotype Analysis	Dr. P.V.V. Gowri Sairam,	5	5	5
15.	MicroRNAs to nanovesicles: Exploration of novel therapeutics for skin diseases	Dr. Gopinath Meenakshisundaram,	5	5	5
16.	Molecular cytogenetics in Genetic Diagnosis	Dr. Jayaram S Kadandale,	5	5	5

17.	**Hands-on session – Module 1: Cytogenetics**				
	1) Culture set-up 2) Harvesting and banding 3) Karyotyping 4) FISH	Ms.Supriya B	5	5	5
		Mr. Manju HC			
18.	**Hands-on session – Module 2: Molecular Biology & Diagnostics**				
	1) DNA isolation from blood	Ms. Anju Srinivas	5	5	5
	2) ARMS PCR for mutation analysis	Ms. Sumaiya Kalaigar			
	3) DNA isolation from bacteria	Mrs. Nishitha R Kumar	5		
	4) 16SrDNA PCR	Mr. Mohammed Kaleem Ullah	5	5	5
19.	**Hands-on session – Module 3: Cell biology**				
	1) Mammalian cell culture 2) Cell counting 3) Subculturing	Mr. Adel Mohammed Saleh	5	5	5
	4) Collection of protein lysates & Western blot	Mr. Anshu Kumar Yadav			
20.	**Hands-on session – Module 4: Gene expression studies**				
	1) RNA isolation & quality check	Ms. Anju Srinivas	5	5	5
	2) cDNA synthesis	Ms. Vaibhavishree M	5	5	5
	3) Quantitative real time PCR	Ms. Deepthi V	5	5	5
	4) Data analysis	Ms. Supriya B	5	5	5
		Mr. Anshu Kumar Yadav	5	5	5

4. Evaluation of the program as a whole:
   a. Did the topics selected cover all important aspects of the specialty? Yes/No
   b. Have you received handouts of all the sessions? Yes/No
   c. Did the sessions help in improvement of your understanding? Yes/No
   d. Was there adequate time for discussion? Yes/No
   e. Do you think there is need for regular workshops of this type in future?
      Yes/No If yes, how often? _____
   f. Any more techniques you can suggest which can be included in the workshop
      _____
   g. How did you get the information regarding this workshop?
      i. By circulars to your institution
      ii. Invitation by organizers
      iii. Internet / Institution website/ social media
      iv. Any other

h. How was the arrangements made by the organizers (please tick)

	Excellent	Satisfactory	Not satisfactory
Registration	✓		
Food	✓		
Conduct of workshop	✓		

i. Any problems faced by you as a participant?

No

j. Suggestions for future workshop

Date: 24/06/2022

Name and Signature